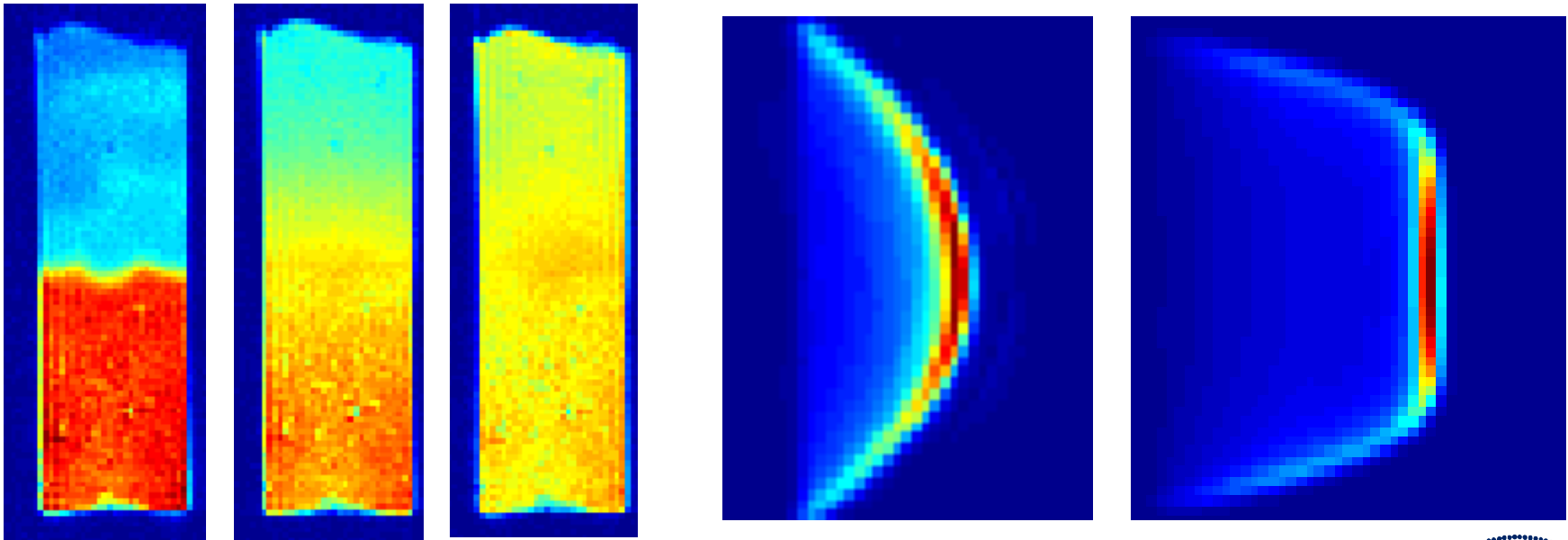


Transport Effects in Cellulose Fiber Beds Using Magnetic Resonance Imaging

R.L. Powell

David M. Lavenson, E.J. Tozzi, M.J. McCarthy

University of California, Davis



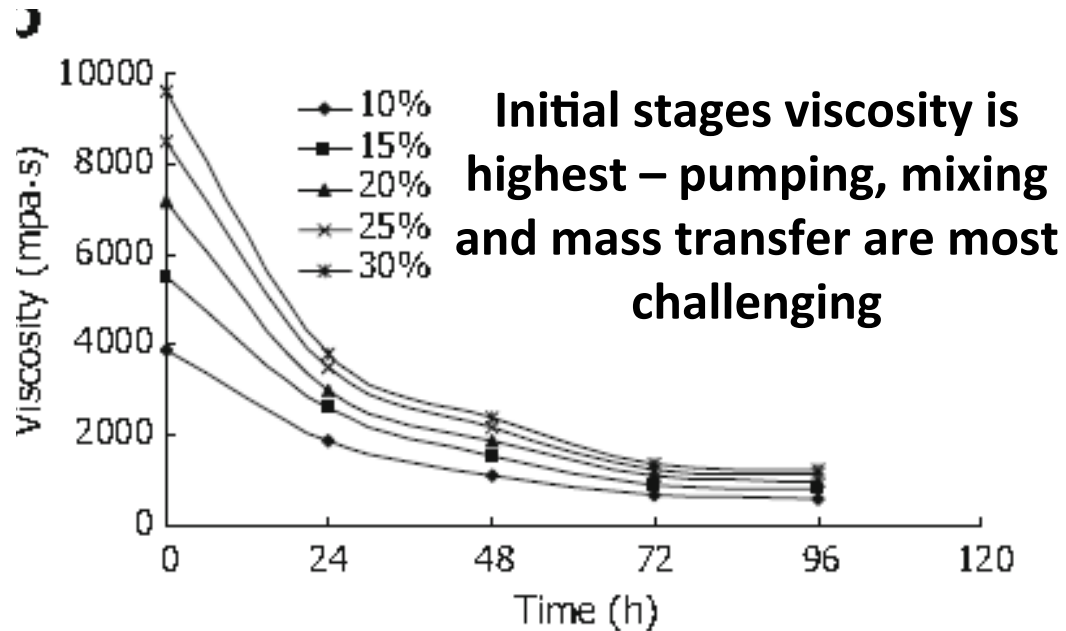
Outline

- **Biomass Processing**
- **MRI**
- **Diffusion & Adsorption**
- **Fluid Mechanics & Rheology**
- **Conclusions**



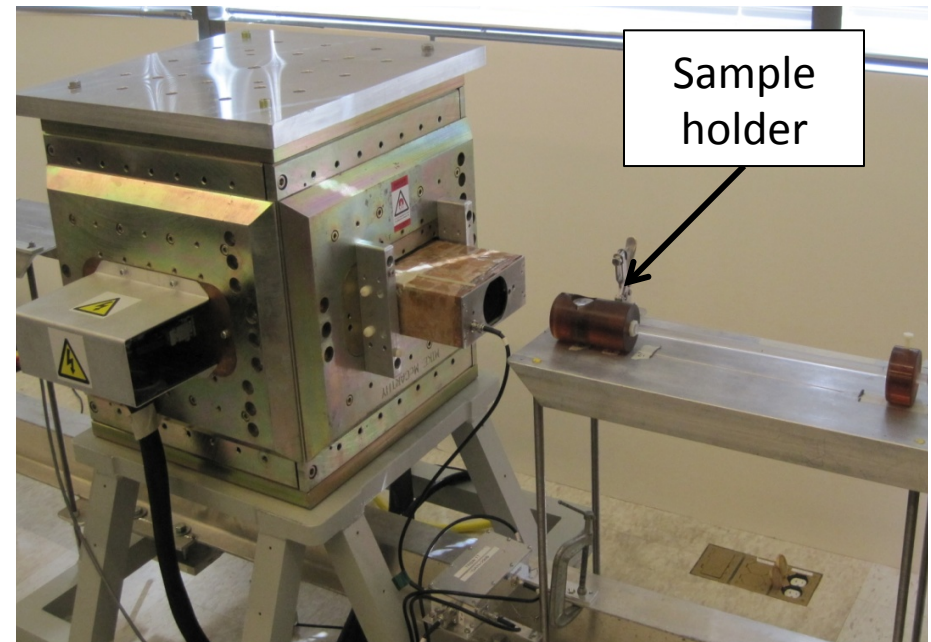
Some Issues

- High cellulose content implies poor mixing of enzyme and cellulose during hydrolysis



Goals

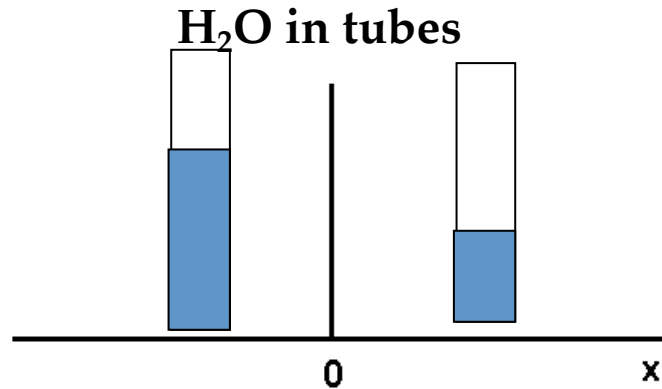
- Use diffusing molecules with similar adsorption behavior and molecular weight as enzymes
- Measure diffusion in various cellulosic fiber beds
- Measure yield stress
- Use magnetic resonance imaging (MRI)



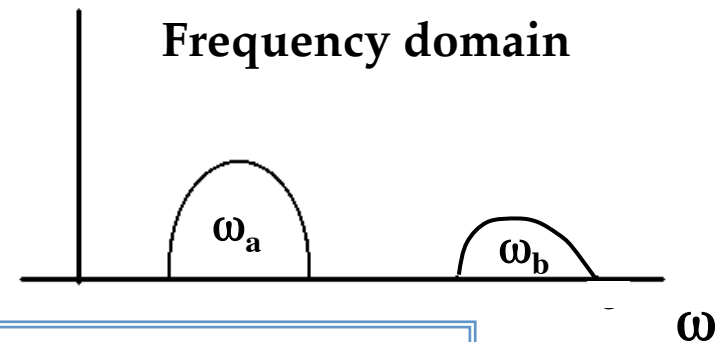
1T Aspect permanent magnet shown above; cartoon of sample holder and magnet below.



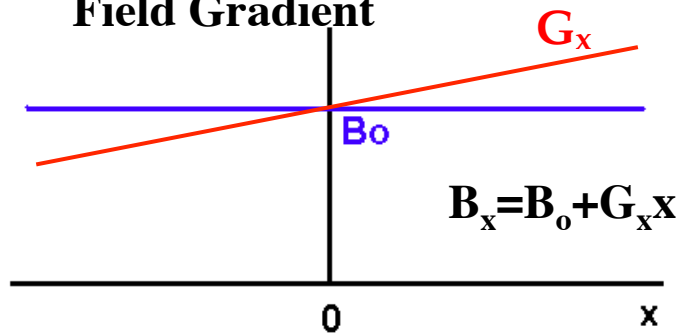
MRI



Transform Frequency into Spatial Information

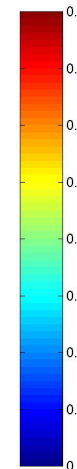
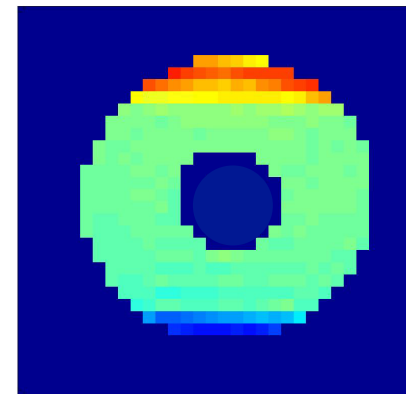


Linear Magnetic Field Gradient

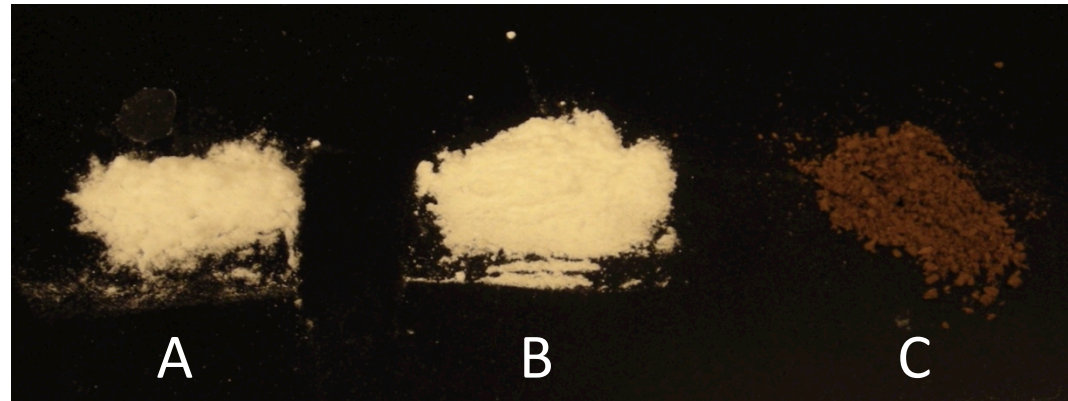


$$\omega = \gamma (B_0 + G_x x)$$

Larmor Relation



Fibers



Fiber type	Length (LW)	Length (NW)	Width (LW)
a) Solka-Floc 200EZ	0.207 mm	0.183 mm	26.4 μm
b) Solka-Floc C100	0.349 mm	0.273 mm	31.7 μm
c) NREL PCS	-	-	-

short 

medium 

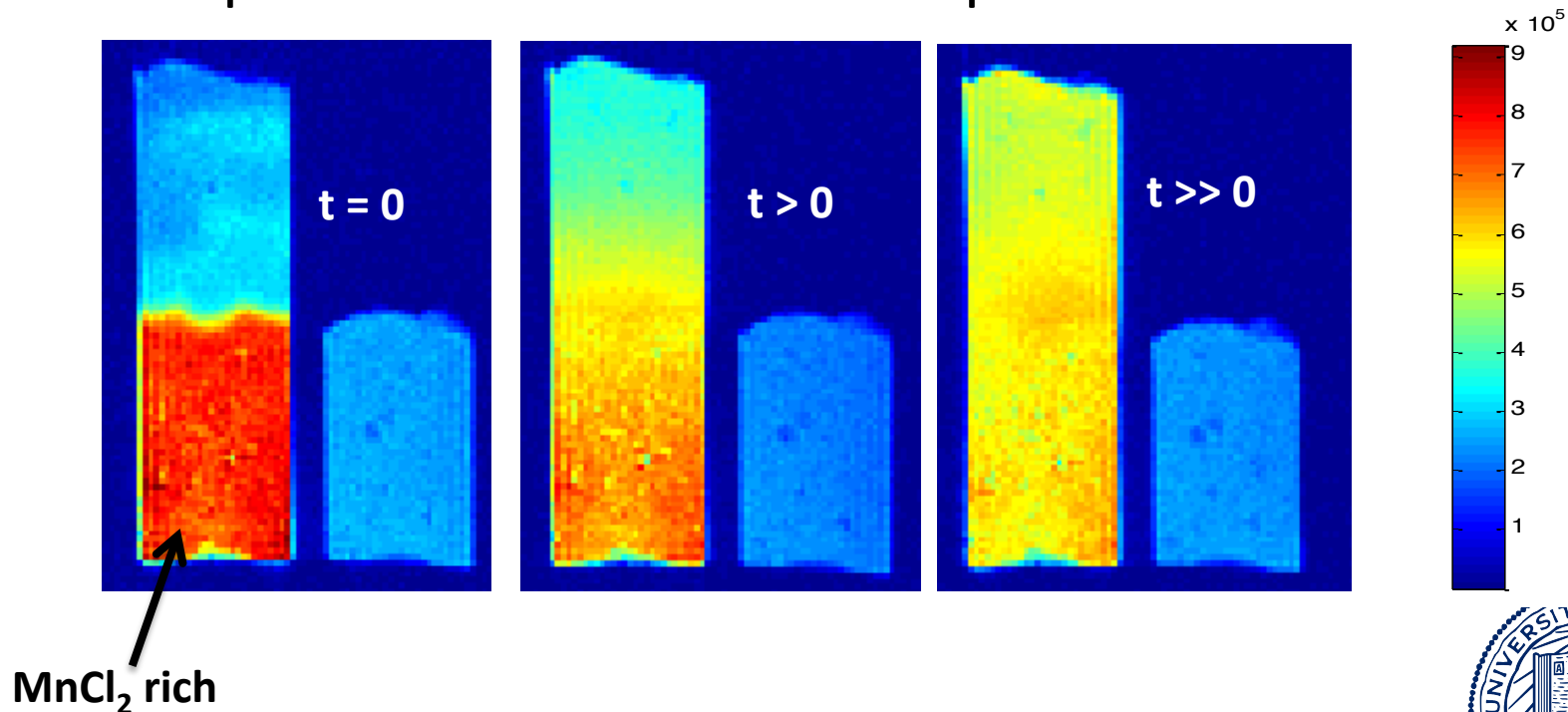
Average length of fibers used
(LW = Length Weighted)

1 mm 

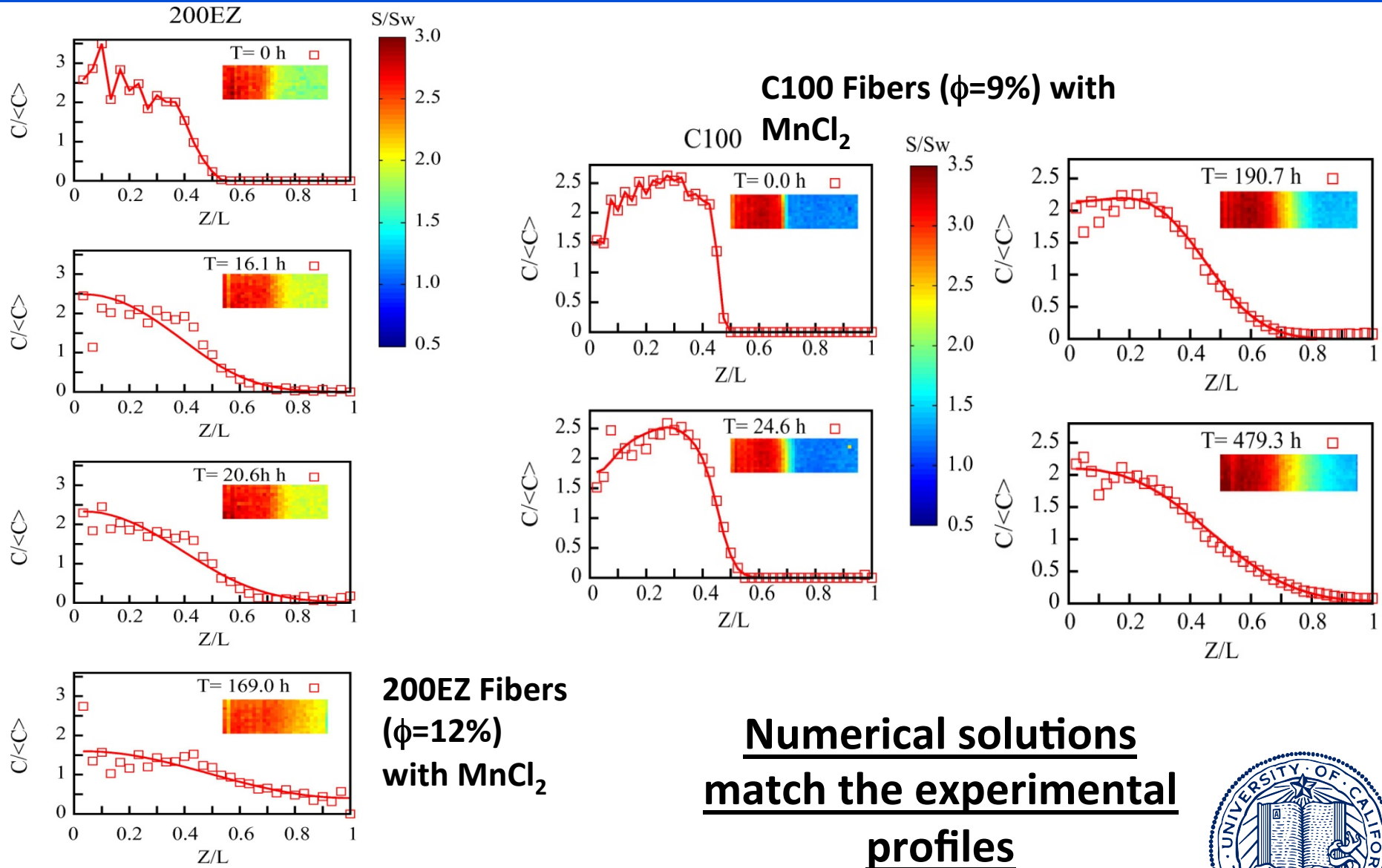


Images and Analysis

- Images taken at various time points
- Concentrations determined from calibration relationship
- Crank-Nicolson finite difference numerical solution used to match experimental and numerical profiles

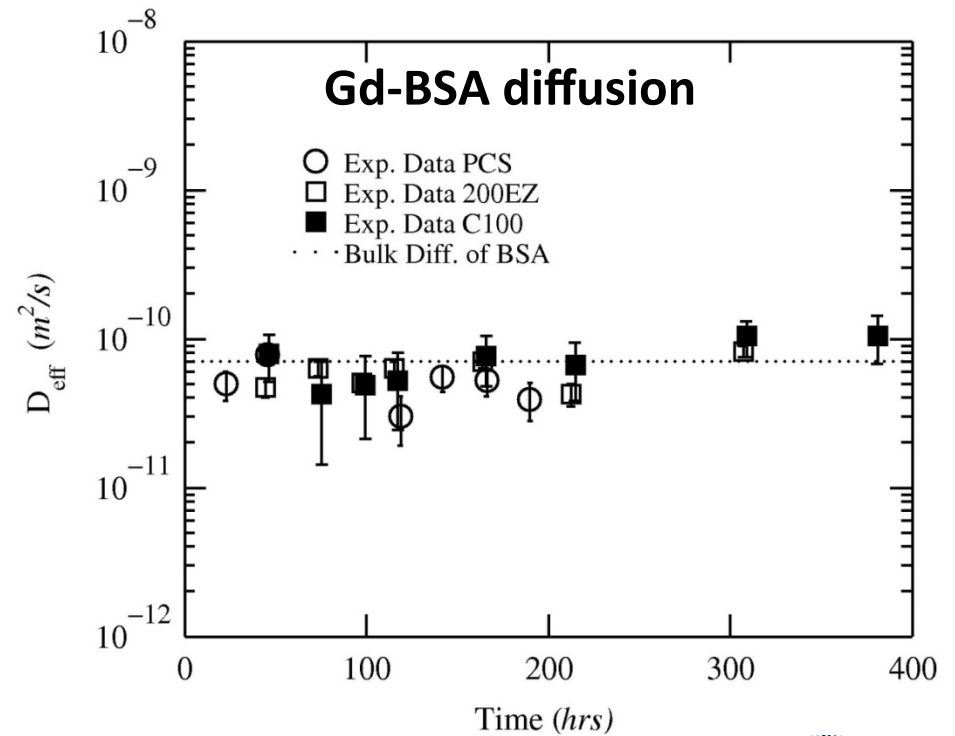
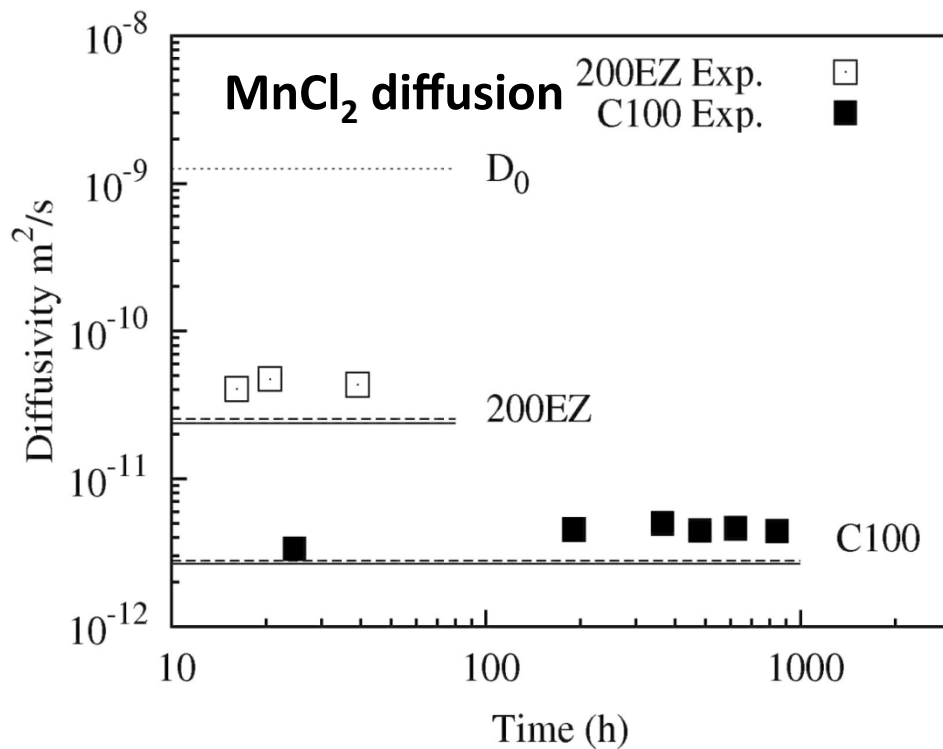


Concentration Profiles



Diffusion Model Summary

Fiber	C100		200EZ		PCS		Bulk, D_0
	Exp.	Model	Exp.	Model	Exp.	Model	
$D_{\text{MnCl}_2}, m^2 s^{-1}$	4.39×10^{-12}	2.79×10^{-12}	4.37×10^{-11}	2.56×10^{-11}	-	-	1.26×10^{-9}
$D_{\text{BSA}}, m^2 s^{-1}$	7.07×10^{-11}	6.10×10^{-11}	5.98×10^{-11}	5.81×10^{-11}	5.05×10^{-11}	5.18×10^{-11}	7.00×10^{-11}



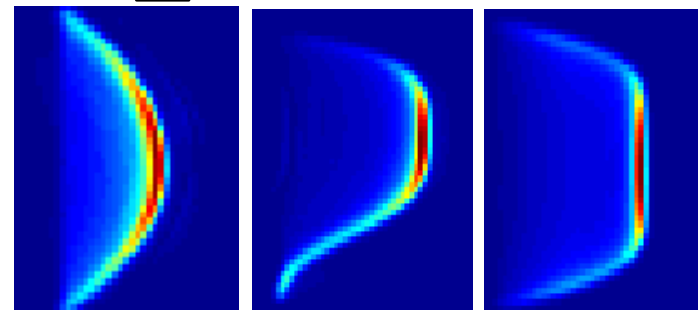
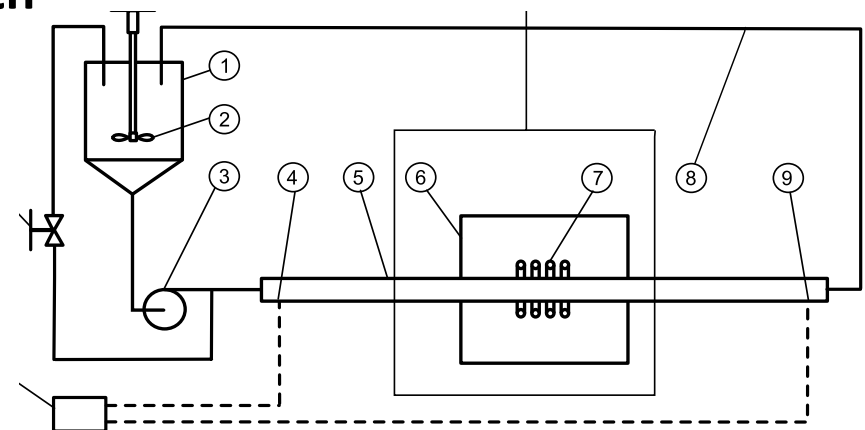
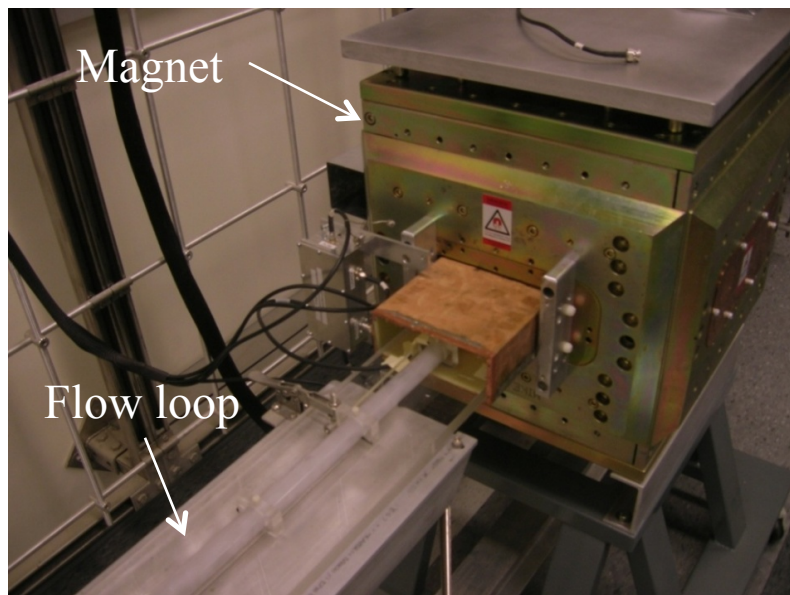
Rheology

- Recent biomass literature
 - Non-Newtonian behavior
 - Shear-thinning and a yield stress
 - Understand rheology of suspensions during biomass digestion (enzymatic hydrolysis)
- MRI Flow Imaging
 - Effective in-line rheometer for rheological measurements

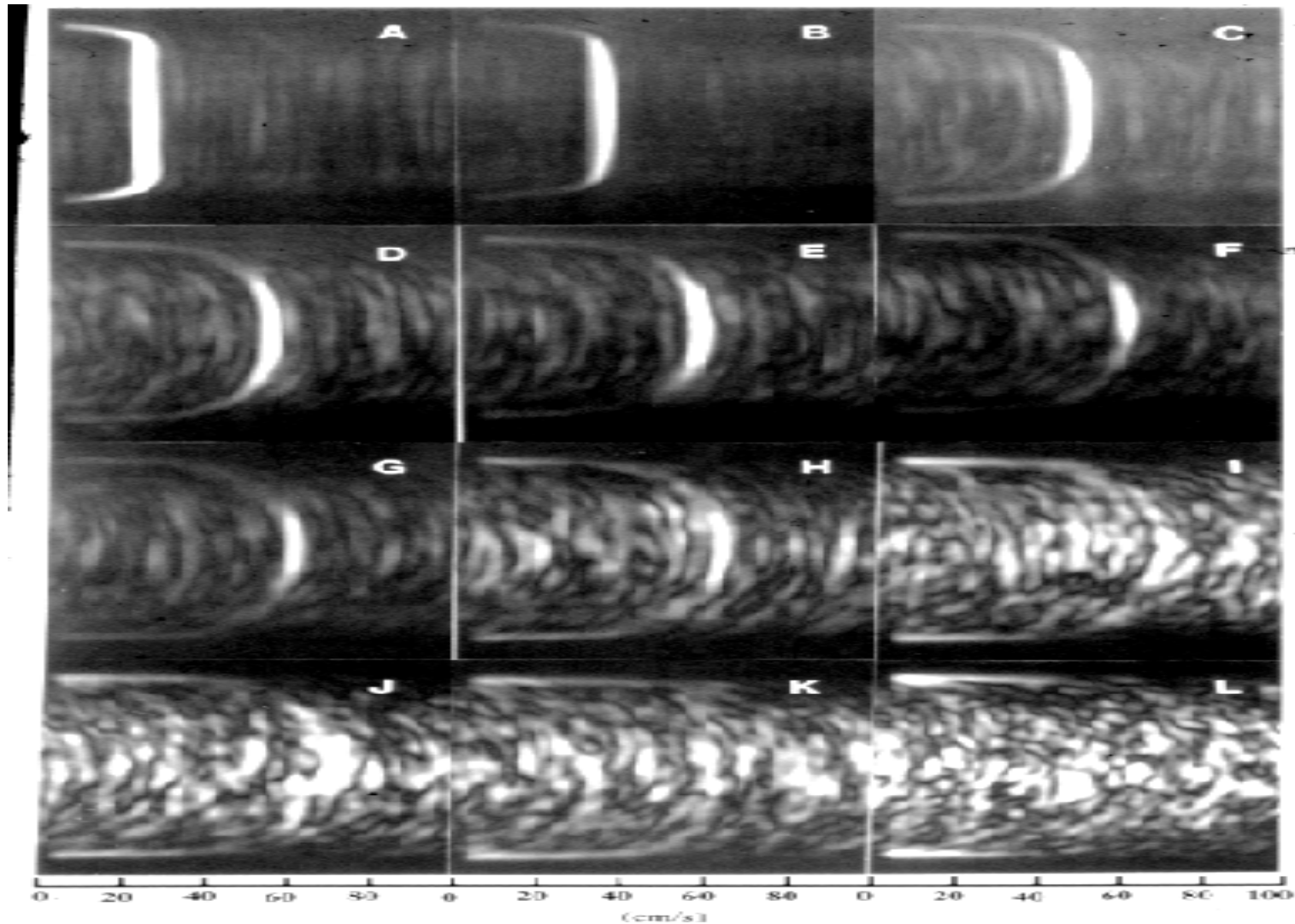


Velocity Measurements via MRI

- **1T Aspect permanent magnet based spectrometer**
 - 30 G/cm peak gradient strength
 - 38x36 mm cylindrical coil
 - 2.08 m long, 19mm diameter pipe
- **Pulsed gradient spin echo sequence with velocity encoding**
- Resolve settling, asymmetry, and plug flow profiles of high solids suspensions
- Simultaneously measure pressure drop across the pipe



0.5% Hardwood Pulp Suspension



Viscosity Measurement

Measure
 ΔP

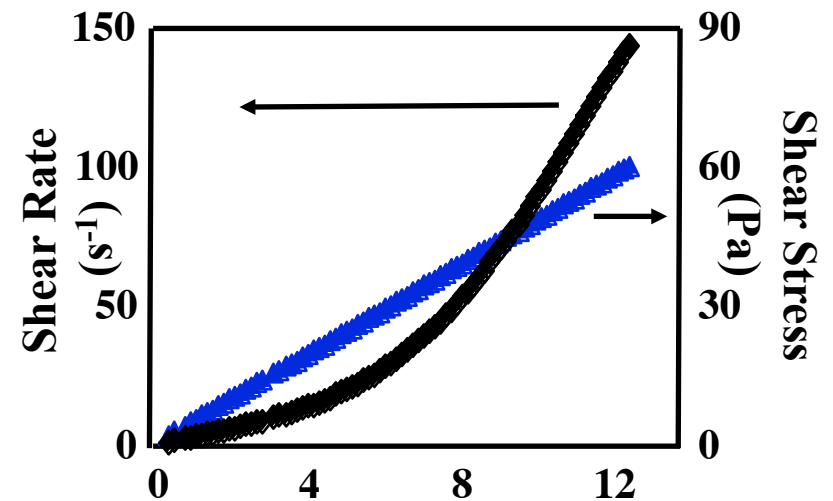
Measure
 $v(r)$

$$\sigma(r) = \frac{\Delta P}{2L} r$$

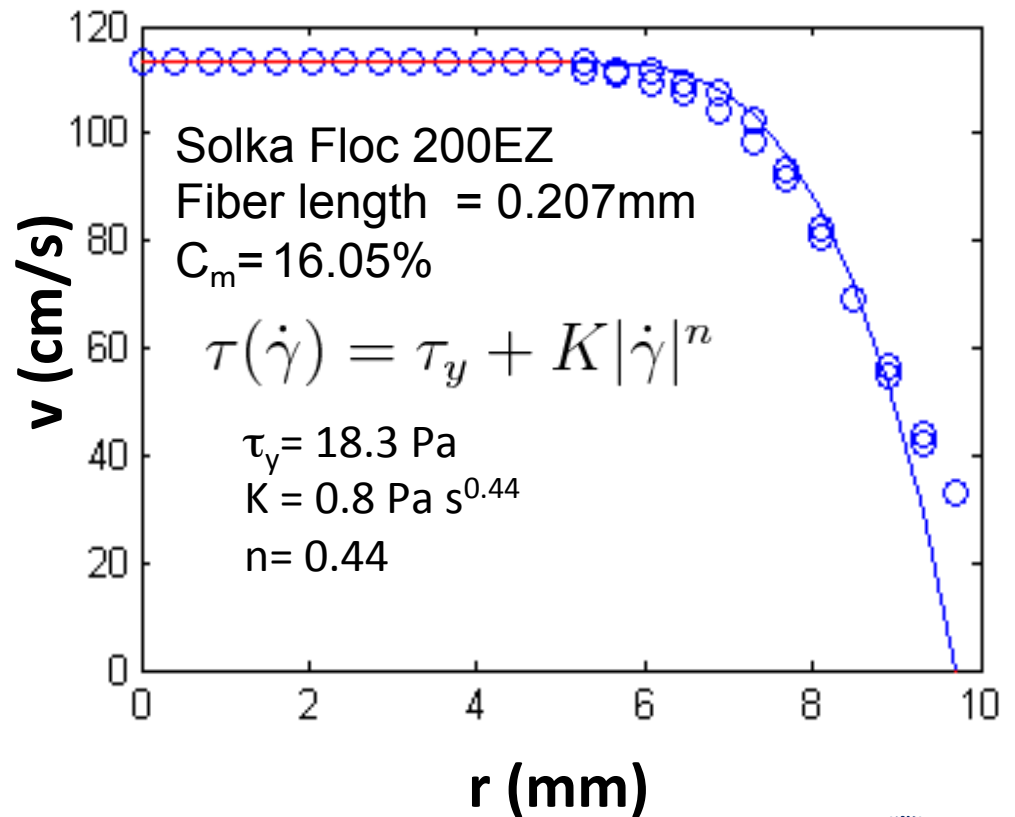
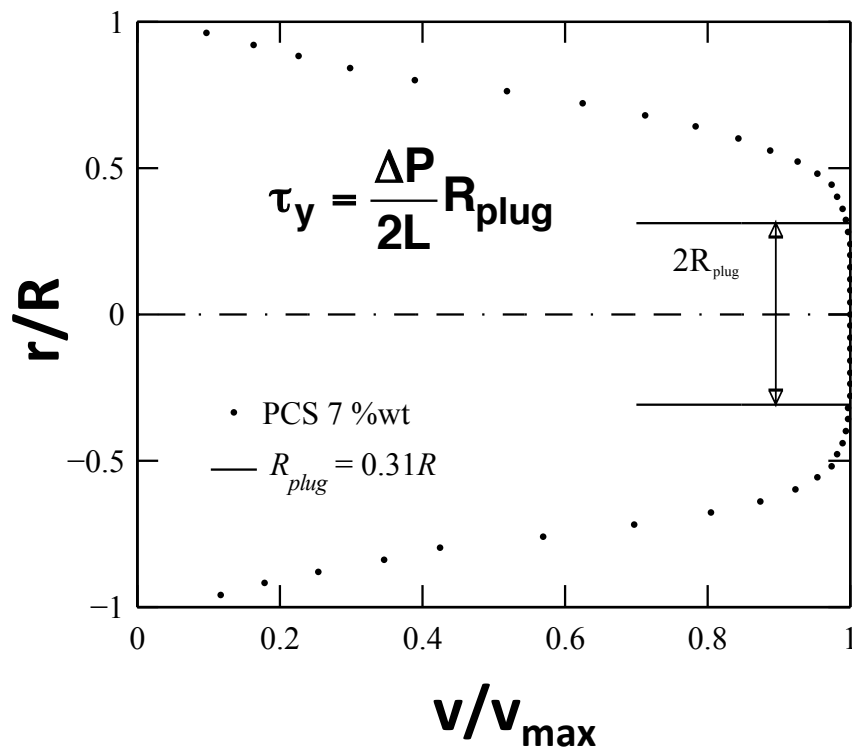
$$\dot{\gamma}(r) = \left| \frac{dv(r)}{dr} \right|$$

Shear Viscosity

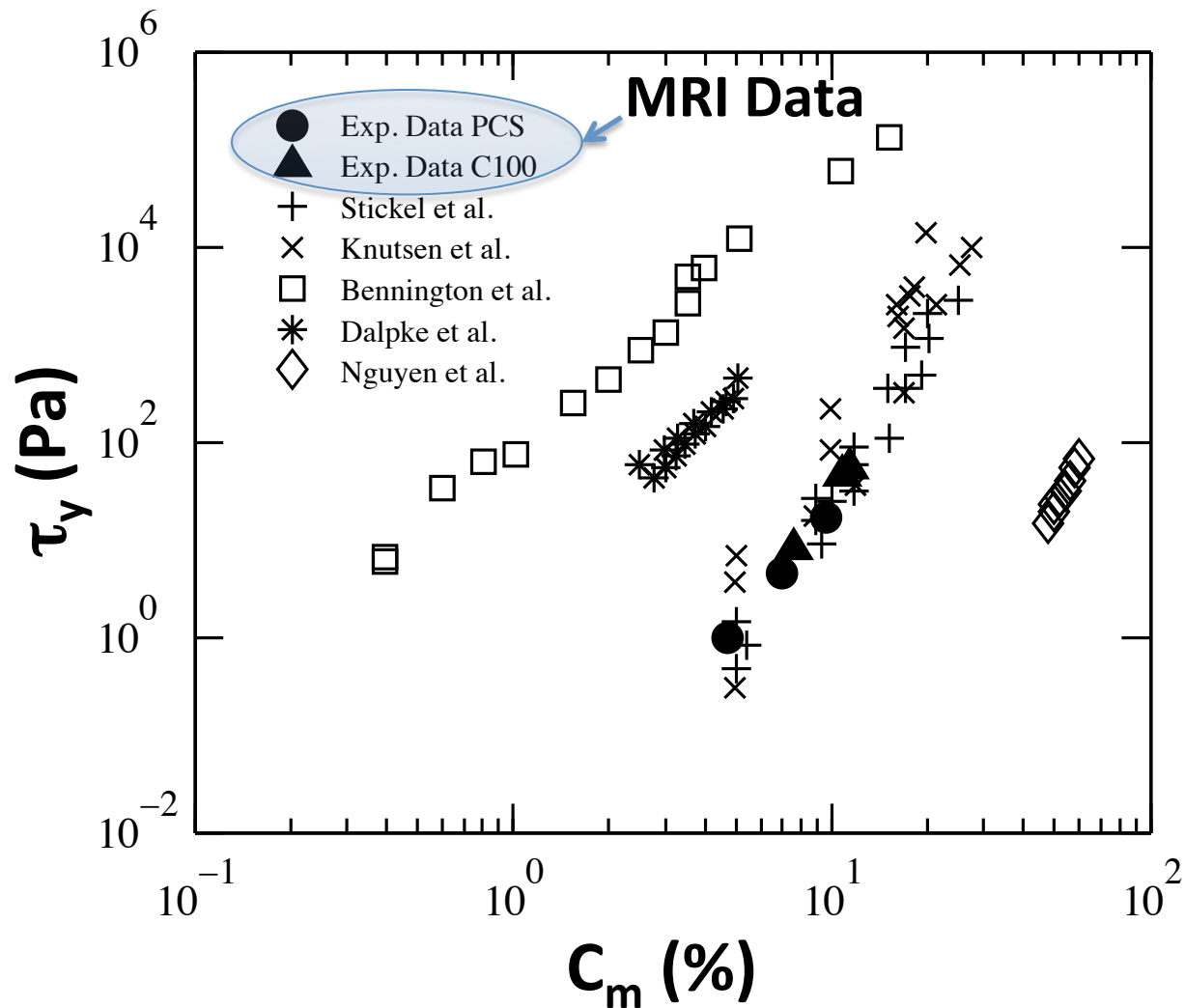
$$\eta(\dot{\gamma}) = \frac{\sigma(\dot{\gamma})}{\dot{\gamma}}$$



Yield Stress



Summary of Results



Summary of Results

Fiber Type	Length Dist. (mm)	σ_0 Range (Pa)	C_m Range (%)	a	b	Reference
Semi-bleached pulp	1.94–2.51	0.3 – 2.0×10^4	0.004 – 0.4	99.7	2.45	(Bennington et al. 1990)
Aspen & Birch pulp	0.94–1.43	70 – 500	2 – 6	2.8	2.93	(Dalpke & Kerekes 2005)
PCS	0.001–2.0	1.0 – 5.0×10^4	5 – 30	1.4×10^{-4}	5.27	(Stickel et al. 2009)
PCS	0.001–3.0	0.1 - 1.0×10^4	5 – 30	3.1×10^{-4}	5.37	(Knutsen & Liberatore 2009)
TiO ₂ (“spherical”)	2×10^{-4}	10-100	50 - 70	1.0×10^{-10}	6.55	(Nguyen et al. 2006)
PCS (MRI)	0.349	1.0 – 20	7 – 15	1.9×10^{-3}	4.00	
C100 (MRI)	0.001-2.0	8.0 - 60	5 - 10	5.0×10^{-4}	4.81	



Conclusions

- Simple diffusion-adsorption model agrees well with experimental results
- Adsorption significantly hinders diffusion
 - Strong dependence on adsorption (MnCl_2)
- No measured adsorption of BSA on cellulose fibers
- Yield stress measured for PCS

